

Page 1 of 4						
*required for saving		Tracking #:				
Facility ID:		*Survey Year:				
Facility Characte	ristics					
*Ownership (checl	k one):					
🗌 For profit	\square Not for profit, including church	□ Government				
☐ Military	Veterans Affairs	\Box Physician owned				
		,				
If facility is a Hos	pital:					
*Number of patien	t days:					
	sions:					
For any Hospital:						
*Is your hospital a	teaching hospital for physicians and/or	r physicians-in-training? Yes No				
If Yes, what	at type: Major	_ Graduate Undergraduate				
*Number of beds s	set up and staffed in the following locat	ion types (as defined by NHSN):				
a. ICU (ind	cluding adult, pediatric, and neonatal le	evels II/III and III):				
b. All othe	r inpatient locations:					
If facility is an An	nbulatory Surgery Center (ASC):	□ No ASC or not operational in this survey year				
Setting: With	nin a hospital Free-standing					
	· ·	ent of procedures that were surgical:%				
		ere discharged or transferred to the following places:				
Home/Custom		%				
	e center (facility other than this one):	%				
-	spital (Emergency or inpatient):	%				
Infection Control	Practices					
*Number of infection	on preventionists (IPs) in facility:					
a. Total hours per week performing surveillance:						
b. Total hours per week for infection control activities other than surveillance: Continued >>						
Assurance of Confidentiality: The information obtained in this surveillance system that would permit identification of any individual or institution is collected with a guarantee that it will be held in strict confidence, will be used only for the purposes stated, and will not otherwise be disclosed or						
released without the consent of the individual, or the institution in accordance with Sections 304, 306 and 308(d) of the Public Health Service Act						
(42 USC 242b, 242k, and 242m(d)).						
Public reporting burden of this collection of information is estimated to average 30 minutes per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of						
information. An agency may not conduct or sponsor, and a person is not required to respond to a collection of information unless it displays a						
	currently valid OMB control number. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to CDC, Reports Clearance Officer, 1600 Clifton Rd., MS D-74, Atlanta, GA 30333, ATTN: PRA (0920-0666).					
		2, 2000 C				
CDC 57.103 (Front) Rev. 6 release 7.1						



Page 2 of 4							
Facility Microbiology Laboratory Practices							
*1. Does your facility have its ow	n laboratory that performs antim	nicrobia	l susceptibility tes	sting?			
🗆 Yes 🛛 No							
If No, where is your facility's a	antimicrobial susceptibility testing	g perfor	med? (check one	2)			
Affiliated medical ce	nter 🗌 Commercial referral	laborat	orv				
			-				
*2. Does the laboratory use CLS	I (formerly NCCLS) antimicrobia	al susce	ptibility standards	\$?			
🗆 Yes 🛛 No							
If Yes, specify the version of t	he M100 document that the lab	oratory	uses: M100- S				
*3. For the following organisms (please indicate which methods a	are usec	l for:				
(1) primary susceptibility	testing and						
(2) secondary, suppleme	ental, or confirmatory testing (if p	performe	ed).				
	ot perform susceptibility testing,	please	indicate the meth	nods used at the referral			
laboratory.							
_	des listed below the table.	(2) 60	oondom.	Comments			
Pathogen	(1) Primary	(2) 50	condary	Comments			
Coagulase-negative staphylococ Staphylococcus aureus		<u> </u>					
Enterococcus spp.		<u> </u>					
Enterobacteriaceae		<u> </u>					
Pseudomonas aeruginosa		<u> </u>					
Acinetobacter spp.		<u> </u>					
Stenotrophomonas maltophilia		<u> </u>					
1 = Kirby-Bauer disk diffusion	5.1 = MicroScan walkaway rapid		10 = E test				
2 = Vitek (Legacy)	5.2 = MicroScan walkaway rapid	ntional		agar screen (BHI + vancomycin)			
2.1 = Vitek (209403)	5.3 = MicroScan auto or touchsca			ibe in Comments section)			
3.1 = BD Phoenix	6 = Other micro-broth dilution met						
4 = Sensititre	7 = Agar dilution method						
*4. Does the laboratory confirm vancomycin-resistant staphylococci using a second method? 🗌 Yes 🗌 No							
If Yes, please indicate methods: (check all that apply)							
🗆 Kirby-Bauer disk diffusion 🛛 MicroScan walkaway rapid 🔹 🗆 E test							
🗌 Vitek (Legacy)	🗌 MicroScan walkaway co	licroScan walkaway conventional Vancomyc		nycin agar screen (BHI + nycin)			
🗌 Vitek 2	☐ MicroScan auto or touc	☐ MicroScan auto or touchscan ☐ Other (spe		specify):			
BD Phoenix	Other micro-broth diluti	□ Other micro-broth dilution method					
□ Sensititre	□ Agar dilution method						
*5. Has your laboratory implemented the revised cephalosporin and monobactam breakpoints for Enterobacteriaceae recommended by CLSI as of 2010?							



Page 3 of 4 Facility Microbiology Laboratory Practices								
*6. Does the laboratory perform a spec		□ No						
If Yes, please indicate what is done if ESBL production is detected: (check one) Change susceptible and intermediate interpretations for third generation cephalosporins and aztreonam to resistant								
\Box Suppress the results for thi	\Box Suppress the results for third generation cephalosporins and aztreonam for the report							
No changes are made in the interpretation of cephalosporins and aztreonam, the test is used for epidemiological or infection control purposes								
*7. Has your laboratory implemented the revised carbapenem breakpoints for Enterobacteriaceae recommended by CLSI as of 2010?								
 *8. Does your laboratory perform a special test for carbapenemase production? Yes No If Yes, please indicate what is done if carbapenemase production is detected: (check one) Change susceptible carbapenem results to resistant Report carbapenem MIC results without an interpretation No changes are made in the interpretation of carbapenems, the test is used for epidemiological or 								
infection control purposes								
*9. Does your laboratory perform colistin or polymyxin B susceptibility testing for drug-resistant gram negative bacilli?								
If Yes, please indicate methods: (c	heck all that apply)							
☐ Kirby-Bauer disk diffusion	🗌 MicroScan walkaway rapid	E test						
□ Vitek (Legacy)	☐ MicroScan walkaway conventional	Vancomycin agar screen (BHI + vancomycin)						
□ Vitek 2	\Box MicroScan auto or touchscan	Other (specify):						
□ BD Phoenix	□ Other micro-broth dilution method							
□ Sensititre	\Box Agar dilution method							
*10. Does your facility have its own laboratory that performs antifungal susceptibility testing for <i>Candida</i> species? Yes No If No, where is your facility's antifungal susceptibility testing performed? (check one) Affiliated medical center Commercial referral laboratory Not offered by my facility								
 11. If antifungal susceptibility testing is performed at your facility or an outside laboratory, what methods are used? (check all that apply) Broth macrodilution Broth microdilution YeastOne colorimetric microdilution 								
	□ Vitek 2 card □ Disk diffusion □ Other (specify):							



Page 4 of 4								
Facility Microbiology Laboratory Practices								
*12. Is antifungal susceptibility testing performed automatically/reflexively for <i>Candida</i> spp. cultured from normally sterile body sites (such as blood), without needing a specific order or request for susceptibility testing from the clinician?								
🗆 Yes	□ Yes □ No							
If Yes, wl	If Yes, what antifungal drugs are tested automatically/reflexively? (check all that apply)							
Flucor	nazole	□ Itraconazole	□ Voriconazole	□ Caspofungin				
🗌 Micafı	ungin	🗌 Anidulafungin	☐ Flucytosine	□ Other				
	*13. What is the primary testing method for <i>C. difficile</i> used most often by your facility's laboratory or the outside laboratory where your facility's testing is performed? (check one)							
🗌 Enzym	\Box Enzyme immunoassay (EIA) for toxin							
🗌 Cell c	ytotoxicity	neutralization assay						
	ic acid amp	plification test (NAAT) (e.	g., PCR, LAMP)					
🗌 Glutar	nate dehyo	drogenase (GDH) antiger	ו plus EIA for toxin (2-st	tep algorithm)				
	plus NAAT	(2-step algorithm)						
\Box GDH plus EIA for toxin, followed by NAAT for discrepant results								
□ Toxigenic culture (<i>C. difficile</i> culture followed by detection of toxins)								
Other (specify):								